

Synthesis and Biological Evaluation of 1,3,4-Triaryl-3-pyrrolin-2-ones, a New Class of Selective Cyclooxygenase-2 Inhibitors

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Abstract—The synthesis and structure–activity relationships (SAR) of a series of novel selective COX-2 inhibitors are reported. The results show that some of the 1,3,4-triaryl-3-pyrrolin-2-ones **1** are more potent as COX-2 inhibitors than celecoxib, and that lactam **1d** has the same selectivity. © 2000 Elsevier Science Ltd. All rights reserved.

The recent discovery of two isoforms of cyclooxygenase, namely COX-1, expressed constitutively in all tissues, and COX-2,¹ expressed during inflammatory processes in affected tissues but not in the gastric mucosa, has prompted extensive research in developing selective COX-2 inhibitors that lack the gastrointestinal and nephrotoxic side effects² of currently available nonsteroidal anti-inflammatory drugs,³ all of which inhibit both COX-1 and COX-2 (e.g., diclofenac, naproxen, indomethacin). Epidemiological studies also suggest that selective COX-2 inhibitors may retard the progression of Alzheimer's disease⁴ and be useful for the chemopreventive treatment of polyposis and cancer.⁵ Three different structural types of highly selective COX-2 inhibitors⁶ have been reported to date: methanesulfoanilides (e.g., nimesulide⁷), methylsulfonyl or sulfonamido substituted tricycles (e.g., rofecoxib,⁸ celecoxib⁹), and analogues based on nonselective inhibitors (e.g., L-761,066)¹⁰ (Fig. 1).

The spatial orientation of the aromatic rings in these structures is known to be critical for selective COX-2 inhibition. Based on this hypothesis, we report the synthesis and structure–activity relationships (SAR) studies of some 1,3,4-triaryl-3-pyrrolin-2-ones **1**, a new class of selective COX-2 inhibitors.

These lactams **1** were studied with different molecular modeling tools. Molecules were built using the module PREP of the AMBER program¹⁰ and manually docked into the active site of COX-2. Complexes were geometrically optimized using the parm91 set of parameters with a dielectric constant of 4R and a 12 Å cutoff to disregard non-bonded interactions. Initial structures of the different complexes were generated using the information gathered from site-directed mutagenesis studies,¹¹ available crystal structures, and ligand molecular electrostatic potentials computed at the Hartree-Fock level with a

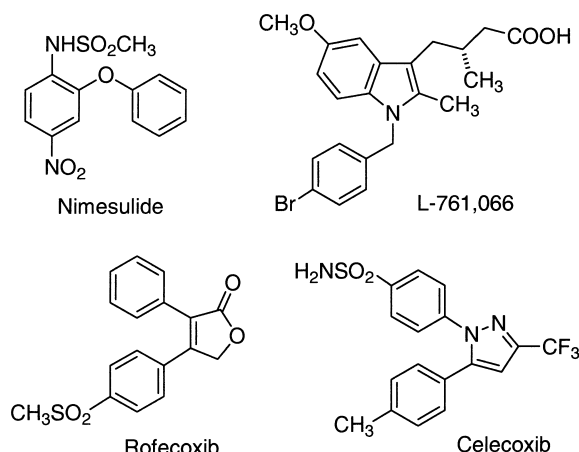


Figure 1.

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6-31G* basis set and GRID¹² calculations on the enzyme active site. All the optimized complexes evidenced good interactions between the sulfone moiety of the ligands and Arg-513. Moreover, the lactam carbonyl group acts as a hydrogen bond acceptor as counterpart of Ser-530. The 3- and *N*-phenyl rings are placed in two hydrophobic pockets composed by the residues Tyr-385, Trp-387, Phe-518, Tyr-348, Val-349, and Leu-352 for the 3-phenyl ring, and Leu-531, Ile-345, Leu-117, and Val-116 for the *N*-phenyl ring (Fig. 2).

The general method employed for the preparation of triaryl-3-pyrrolin-2-ones **1** is outlined in Scheme 1. The synthesis starts from 4-(methylsulfonyl)acetophenone **2**,¹³ which was brominated at the α carbonyl position. Alkylation of appropriate anilines with the resulting α -bromo ketone **3** in ethanol gave amino ketones **4**, which in turn were acylated with a variety of substituted phenyl-

acetyl chloride derivatives. The closure of the lactam ring was satisfactorily accomplished in high yield by an aldol-type cyclization by treatment of amides **5** with DBU. A subsequent dehydration of the resulting lactam alcohols **6** with a *p*-toluenesulfonic acid in refluxing benzene led to the target unsaturated triaryl lactams **1**.

Numerous sites of modification of the lactam ring substituents were explored. A variety of substituents at the *para* position of the *N*- and 3-phenyl rings were investigated in order to provide compounds with good COX-2 activity ($<1 \mu\text{M}$) and acceptable selectivity (ratio $\text{IC}_{50} \text{ COX-1}/\text{IC}_{50} \text{ COX-2} > 10$). In only a few cases we also evaluated the effect of F or Cl atoms at the *ortho* and *meta* positions. No changes were made in the methylsulfone moiety. The IC_{50} values of COX-2 and COX-1, and the selectivity ratios are shown in Table 1. A representative COX-2 inhibitor such as celecoxib and the nonselective

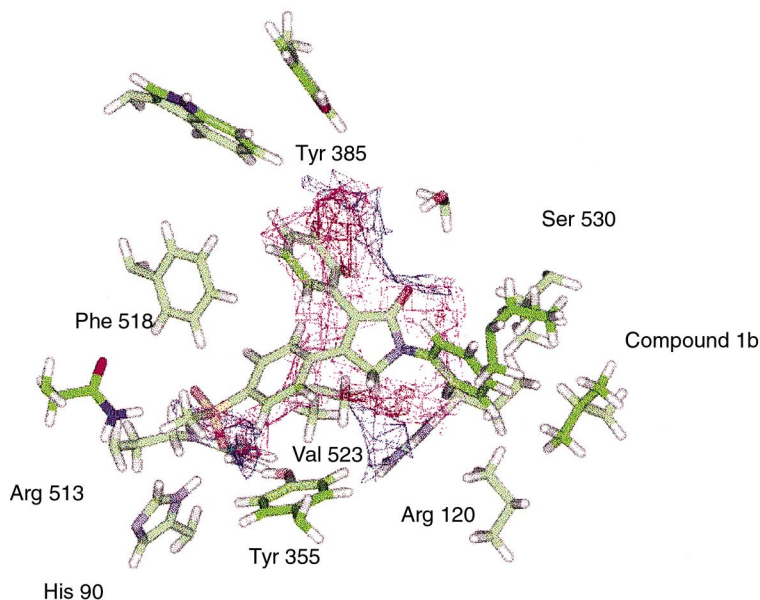
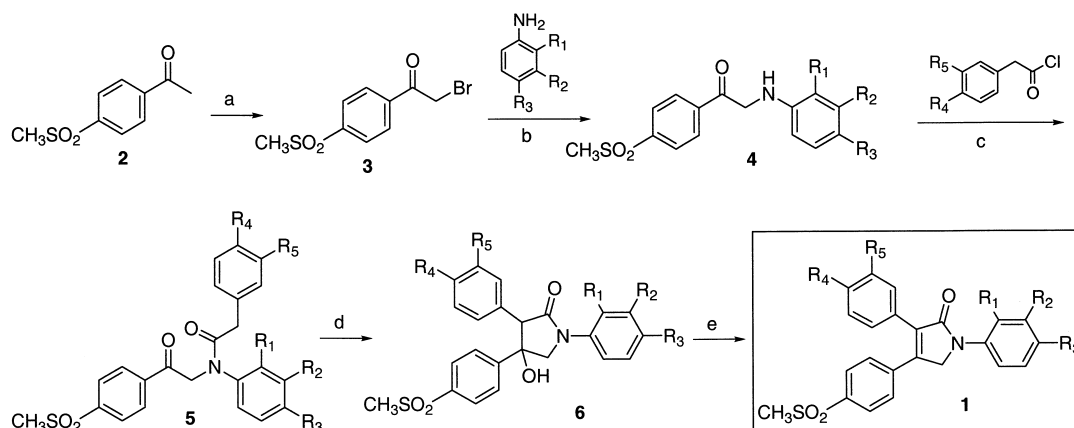


Figure 2. Compound **1b** docked at the COX-2 active site. Color surfaces indicate important regions of interaction at the active site for different probes computed with the GRID program: aromatic (magenta), sulfone (blue), and carbonyl (red).



Scheme 1. (a) Br_2 , CH_2Cl_2 , AcOH , rt, 73%; (b) NaHCO_3 , EtOH , rt, 86–95%; (c) THF , rt, 67–84%; (d) DBU, CH_3CN , 0°C ; (e) *p*-TsOH, C_6H_6 , reflux, 79–92% from **5**.

Table 1. In vitro COX-1 and COX-2 enzyme data for substituted 1,3,4-triaryl-3-pyrrolin-2-ones **1**

Compound	R ₁	R ₂	R ₃	R ₄	R ₅	COX-2 ^a	COX-1 ^a	Selectivity
1a	H	H	H	H	H	0.28±0.09	2.40±0.60	8.6
1b	F	H	H	H	H	0.90±0.14	10.80±1.96	12.0
1c	H	Cl	H	H	H	1.00±0.12	9.80±0.90	9.8
1d	H	H	F	H	H	0.35±0.14	5.86±0.08	16.7
1e	H	H	CH ₃	H	H	1.00±0.09	6.90±1.10	6.9
1f	H	H	CF ₃	H	H	0.98±0.21	5.10±1.20	5.2
1g	H	H	H	CH ₃ O	H	0.34±0.09	2.68±0.85	7.9
1h	H	H	H	F	H	0.31±0.12	2.70±0.61	8.7
1i	F	H	H	F	H	0.32±0.14	2.50±0.48	7.8
1j	Cl	H	H	CH ₃ O	H	0.22±0.05	2.30±0.90	10.5
1k	CH ₃ O	H	H	F	H	1.05±0.25	4.00±1.05	3.8
1l	H	F	H	CH ₃ O	H	0.28±0.10	2.50±0.65	8.9
1m	H	H	CH ₃	F	H	1.05±0.12	3.70±0.59	3.5
1n	H	H	CH ₃ O	F	H	1.20±0.41	5.28±0.94	4.4
1o	H	H	CF ₃	F	H	0.95±0.63	7.20±1.10	7.6
1p	H	H	F	CH ₃ O	H	0.30±0.14	2.50±0.39	8.3
1q	H	H	H	F	F	1.00±0.21	8.20±1.60	8.2
Celecoxib						0.53±0.18	9.73±2.02	18.4
Indomethacin						0.41±0.07	0.19±0.06	0.5

^aIC₅₀ (μM). Average of 2–4 determinations. PGE₂ levels in lipopolysaccharide (LPS)-challenged human whole blood and thromboxane 2 (TxB₂) levels following blood coagulation were measured as biochemical index for COX-2 and COX-1, respectively (see ref 14).

COX inhibitor indomethacin are included for the purpose of a clear SAR summary. As can be seen in the Table, the in vitro COX-2 potency of both the parent lactam **1a** and other substituted lactams bearing a F atom at the *para* position of either the *N*- or 3-phenyl ring (**1d** and **1h**, respectively) or a *para* methoxy 3-phenyl substituent (**1g**) is higher than that of celecoxib. This good COX-2 potency is maintained when there are additional halogen (F or Cl) substituents in the *N*-phenyl ring (**1i**, **1j**, **1l** or **1p**), but strongly decreases with the introduction of bulkier *ortho* or *para* substituents (CH₃, CF₃ or CH₃O). As far as the selectivity ratio COX-1/COX-2 is concerned, most of the lactams **1** proved to be selective COX-2 inhibitors. The best ratios were obtained with lactams **1b** and, especially, **1d**, which bear a fluorine atom at the 2- or 4-position, respectively, of the *N*-phenyl ring. An additional substitution on this ring or at the 3- or 4-position of the 3-phenyl ring does not improve the selectivity.

In summary, we have designed and synthesized a variety of novel 1,3,4-triaryl-3-pyrrolin-2-ones **1** as selective COX-2 inhibitors. These compounds, in particular **1b** and **1d**,¹⁵ may represent a new generation of NSAIDs, useful for symptomatic treatment of inflammatory diseases such as rheumatoid arthritis or osteoarthritis.

Acknowledgements

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15. Spectroscopic data for **1b**: ^1H NMR (300 MHz, CDCl_3) δ 3.08 (s, 3H), 4.83 (s, 2H), 7.24 (m, 3H), 7.40 (m, 5H), 7.54 (d, 2H, $J=8.7$ Hz), 7.74 (td, 1H, $J=1.8, 8.2$ Hz), 7.90 (d, 2H, $J=8.7$ Hz); ^{13}C NMR (75 MHz, CDCl_3) δ 44.3, 54.0, 116.6 (d,

$J=20$ Hz), 124.6, 127.5, 127.8, 128.0, 128.1, 128.6, 128.7, 128.9, 129.5, 130.0, 134.9, 138.2, 140.8, 145.9, 156.5 (d, $J=247$ Hz), 169.0. Spectroscopic data for **1d**: ^1H NMR (300 MHz, CDCl_3) δ 3.08 (s, 3H), 4.77 (s, 2H), 7.13 (t, 2H, $J=9.0$ Hz), 7.41 (s, 5H), 7.53 (d, 2H, $J=8.7$ Hz), 7.80 (dd, 2H, $J=4.8, 9.0$ Hz), 7.89 (d, 2H, $J=8.7$ Hz); ^{13}C NMR (75 MHz, CDCl_3) δ 44.2, 52.6, 115.6 (d, $J=22$ Hz), 120.5, 127.6, 128.4, 128.6, 128.8, 129.2, 130.4, 134.8, 135.7, 137.8, 140.7, 144.2, 159.2 (d, $J=242$ Hz), 168.3.